

OVERTURE IN DEVELOPMENT, PROPERTIES AND CLINICAL ASPECTS OF BIOSURFACTANTS: AN REVIEW

Ashwini H. Bijewar¹, Wrushali A. Panchale^{1,*}, Jagdish V. Manwar², Ravindra L. Bakal¹

¹ IBSS's Dr. Rajendra Gode Institute of Pharmacy, Mardi Road, Amravati-444 602, MS, India.

² IBSS's Dr. Rajendra Gode College of Pharmacy, Mardi Road, Amravati-444 602, MS, India.

*Corresponding Author: Email: wpanchale@gmail.com

Received: 18 April 2021/ Revised: 26 May 2021 / Accepted: 28 June 2021 / Available online: 30 June 2021

ABSTRACT

Biosurfactants are a basically assorted gathering of surface-dynamic substances created by microorganisms. They are amphiphiles, they comprise of two sections- a polar (hydrophilic) moiety and non-polar (hydrophobic) congregation. In spite of a colossal measure of examination work over the most recent twenty years on conserving the creation of biosurfactants, their business accomplishment when contrasted with their manufactured partners actually stays a financial test. Utilization of immobilized organic entity, utilization of NPs, strong state aging, coordinated aging, froth fractionation, and fill and draw method of activity could end up being other promising cycles for the upgraded modern creation of different biosurfactants. Utilization of natural, fortified waste substrates and biosurfactants coproduction with another modern efficient item should be all the more basically concentrated particularly in huge aging vessels. In present work, we have covered various physicochemical and clinical aspects of biosurfactants.

Keywords – Biosurfactants; Nanoparticles; Biofilm; Application.

1. INTRODUCTION

Biosurfactants are produced by extensive variation of diverse microorganisms and possess structures of different chemical and surface properties. They are surface-dynamic substances created by microorganisms comprising of two sections- a polar (hydrophilic) moiety and non-polar (hydrophobic) gathering. A hydrophilic gathering comprises of mono-, oligo- or polysaccharides, peptides or proteins and a hydrophobic moiety as a rule contains soaked, unsaturated what's more, hydroxylated unsaturated fats or greasy alcohols [1-2]. A trademark highlight of biosurfactants is a hydrophilic-lipophilic equilibrium (HLB) which indicates the part of hydrophilic and hydrophobic constituents in surface-dynamic substances [3].

Due to their amphiphilic structure, biosurfactants increment the surface zone of hydrophobic water-insoluble substances, increment the water bioavailability of such substances and change the properties of the bacterial cell surface [4]. Surface action makes surfactants astounding emulsifiers, frothing and scattering specialists (Fig. 1).

In contrast with their synthetically orchestrated reciprocals, they have numerous preferences. They are harmless to the ecosystem, biodegradable, less poisonous and non-perilous [5]. They have better frothing properties and higher selectivity. They are dynamic at extraordinary temperatures, pH and saltiness also, and can be created from mechanical squanders and from results. This last highlight makes modest creation of biosurfactants conceivable and permits using waste substrates and

decreasing their contaminating impact simultaneously [6-8]. They are broadly utilized in numerous ventures, for example, farming, food creation, science, beautifiers and pharmaceuticals. The instances of biosurfactants applications are recorded in many audit papers [9-10].

Exceptional consideration is paid to the utilization of biosurfactants in various parts of ecological biotechnology. Numerous properties of microbial surface dynamic mixes, for example, emulsification/de-emulsification, scattering, frothing, wetting and covering make them valuable in physico, synthetic and natural remediation advancements of both natural also, metal pollutants [11-12]. Biosurfactants increment the bioavailability of hydrocarbon coming about in improved development and corruption of impurities by hydrocarbon-debasing microorganisms present in dirtied soil. In hefty metal dirtied soils biosurfactants structure edifices with metals at the dirt interface, which is trailed by desorption of the metal and expulsion from the dirt surface prompting the increment of metal particles fixation and their bioavailability in the dirt arrangement [13]. The new methodology is the utilization of substantial metal-safe bacterial strains equipped for delivering biosurfactants for expanding the metal-eliminating productivity by phytoremediation [14].

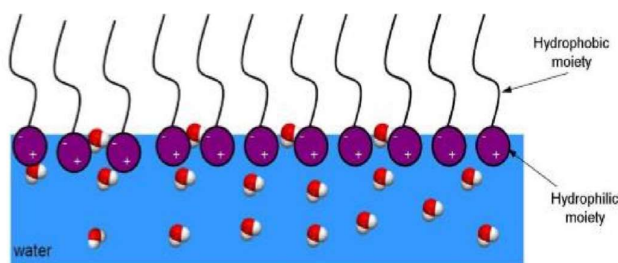


Fig. 1: Biosurfactants, surface tension and formation of micelles

2. SOURCES

Biosurfactants are produced by extensive variation of diverse microorganisms and possess structures of different chemical and surface properties. Microorganisms are proficient in producing different kinds of Biosurfactants such as Pseudomonas, Acinetobacter, Bacillus, Brevibacterium, Clostridium, Rhodococcus, Thiobacillus, Leuconostoc, Citrobacter, Candida, Corynebacterium, Penicillium, Ustilago Aspergillus, Saccharomyces, Enterobacter, and Lactobacillus [15-21].

Penibacillus sp. D9 is a good biosurfactants producer. The various sorts of BioS incorporate glycolipids (mannosyl erythritol, rhamnolipids, sophorolipids, xylolipid, cellobiose lipids trehalose lipids), lipopeptides (subtilisin, vixcosin, serrawetin, surfactin, polymyxin, iturin), polysaccharide-protein edifices, flavolipid, phospholipids, fatty acids, polymeric surfactants (liposan, alasan, emulsan) and lipids [22-25].

The most oftentimes delivered low atomic weight surface dynamic mixes are glycolipids and lipopeptides. The other gathering which has regularly been utilized substitutivity with Biosurfactants to address biomolecules that are surface active are referred to as bioemulsifiers. Bioemulsifiers are surface-dynamic yet don't basically diminish surface pressure, in any case, give consistent emulsions between water blends and hydrocarbons (fluids) [26-27].

3. CLASSIFICATION

Biosurfactants are normal items got from microorganisms, yeasts or parasites. The mind-boggling substance structures and actual properties of biosurfactants for the most part bring about properties equivalent to or surpassing numerous engineered surfactants. Biosurfactants show low poisonousness to freshwater, marine and earthbound biological systems and are likely possibility for an assortment of natural applications. Exploration has to a great extent been centered around the upgrade of oil biodegradation and microbial-improved oil recuperation [28-30] (**Table 1**).

Table 1: The classification of biosurfactants

Class	Biosurfactants	Source microorganism	Mechanism
Glycolipids	Rhamnolipid	<i>Pseudomonas aeruginosa</i> , <i>Pseudomonas sp.</i>	Enhancement of degradation and dispersion of the different classes of hydrocarbon; emulsification of hydrocarbon and vegetable oils; removal of metals form soil.
	Trehalolipid	<i>M. tuberculosis</i> , <i>R. erythropolis</i> , <i>arthrobactor sp.</i> , <i>Nacrodinr sp.</i>	Enhancement in bioavailability hydrocarbons.
	Soporlipid	<i>Torulopsis bombicoia</i> , <i>Torulopsis perophithem</i> , <i>Torulopsis aplicola</i>	Recovery of hydrocarbons form dregs and muds; removal of heavy metals form sediments; enhancement of oil recovery.
Fatty acids, phospholipids and natural lipids	Corynomycolic Acid	<i>Corynebacterium lepus</i>	Enhancement of bitumen recovery.
	Spiculisporic Acid	<i>Penicilinom spiclisporum</i>	Removal of metal ion form aq. Solution; dispersion action for hydrophilic pigments.
	Phospharidylethanolamine	<i>Acinetobacter sp.</i> , <i>a. erythropolis</i>	Increasing the tolerance of bacterial to heavy metal.
Lipopeptides	Surfactin	<i>Bacillus subtilis</i>	Enhancement of biodegradation of hydrocarbon.
	Lichenysin	<i>Bacillus licheinformis</i>	Enhance the oil recovery.
Polymeric biosurfactant	Emulsan	<i>A. calcoacencus RAG-1</i>	Satbilazation of hydrocarbon in water emulsion.
	Alasen	<i>A. radiatoracistens Ka-53</i>	Satbilazation of hydrocarbon in water emulsion.
	Biodispersion	<i>A. calcoacetious A2</i>	Dispersion in limestone in water.

4. PHARMACEUTICAL APPLICATION

4.1. Nanoparticle

Nanoparticle-based therapeutics have been considered as the absolute most encouraging stages in medication conveyance applications because of their capacity to build drug aggregation in strong tumours by improved porousness and maintenance (EPR) and MDR inversion through by passing or hindering group movement [31]. Nanoparticle-interceded cell demise happens by means of *C. albicans* film blasting followed by overflowing out of proteins a d intracellular material. Notwithstanding working as a cyclic lipopetide, the biosurfactants, SUR, has been found to show adaptable bioactive highlights including adjuvant for inoculation and antitumor properties. In light of its special amphipathic properties, SUR has the potential for self-get together (under specific conditions) into nanoparticles to work as a medication transporter for stacking hydrophobic medications [32].

4.2. Inhibition of biofilm formation

Probably the most encouraging possibility for the restraint of biofilms have come from biosurfactants since they have solid antiadhesive, antimicrobial and biofilm interruption properties .It has been suggested that biosurfactants assume a significant part in life forms that produce them by halfway disturbing the developing biofilm and keeping up channels for gas and supplement dissemination and it is consequently to be expected that they are powerful in upsetting biofilms at proper fixations. Analysts here highlight the dispersal of a biofilm of pathogenic microbes by diminishing bacterial cell reasonability and the decrease in bacterial bond properties as proof of the compelling of biosurfactants [33-34].

The recommended instrument of act particle might be identified with the official of the biosurfactants atoms to cell divider segments or the cell surface bringing about extreme changes in external film hydrophobicity. The addition of biosurfactants into the bilayer construction of cell film may bring about the interruption of its respectability. The consequences for both Gram-

negative and Gram-positive microorganisms might be because of the arrival of LPS atoms from the external film or because of the development of transmembrane pores bringing about expanded porousness of the phone divider [35].

5. PHARMACOLOGICAL APPLICATIONS

5.1. In cancer treatment

LPs, glycolipids and different kinds of biosurfactants planting to their primary curiosity and assorted biophysical properties have arisen as conceivable expansive range specialists for malignant growth chemotherapy/biotherapy and as protected vehicles or fixings in medication conveyance definitions. The LPs and SLs are the biosurfactants generally concentrated regarding anticancer potential. The LPs are made out of a peptide and an unsaturated fat chain and have been appeared to show antitumor action. In vitro Reports on the LPs, specifically SUR, Iturin and Fen Bacillus gycin, propose that they have antitumor exercises. Iturin has been appeared to inhibit the expansion of M Da-MB-231 malignancy cells [36].

The anticancer systems of LPs have been widely considered and SUR Bacillus has been found to show an anti-proliferative impact through apoptosis enlistment, cell cycle capture and endurance flagging concealment [37]. Among the proposed employments of SLs are their potential in human cervical malignancy treatment [38]. In remedial and protection xerograph models of B16-EGFRvIII melanoma cells, the self-adjuvant LP antibody micelles viably forestalled tumours development just as tumorigenesis. Different anticancer mechanisms for SLs have been proposed including a role in differentiation and apoptosis [39] (Table 2).

Table 2: Various Biosurfactants used in Anticancer therapy

Class	Biosurfactant Name	Source	Effect on cell line
Lipopeptide	Surfacin	<i>Bacillus subtilis</i>	Suppression of LOVO(colon carcinoma)
Lipopeptide	Surfacin	Bacillus natto TK-1	Killing of MCF-7(breast cell)
Lipopeptide	Iturin	Bacillus subtilis	Inhibition of K562 leukemia cell
Glycolipid	Mannosylerythritol lipid-A, Mannosylerythritol lipid-B	Candida Antarctica T-34	Induced HL60 (leukemia cell line)
Sophorolipid	Sophorolipid	Candida bombicola ATCC 22214	Increase in LN-229
Sophorolipid	Di-acetylated lactonic C18:1	Wickerhamiell domercqiae	Apoptosis in liver cell (H7402)
Sophorolipid	Various derivatives	Candida bombicola ATCC 22214	Antiproliferation of HeLa cell
		Candida bombicola ATCC 22214	Killing of human pancreatic cell
		Wickerhamiell domercqiae	Inhibiting of oesophageal cancer cell
		Starmerella bombicola	Killing of MDA-MB-231 breast cancer cell

5.2. In wound healing

A wide variety of bioactive metabolites, including biosurfactants, are seen as having potential for dermatological applications including wound recuperating. In vitro the injury recuperating capability of SPB1 LP on B. subtilis extraction wounds prompted in trial rodents. They found a significant increment in the level of wound conclusion contrasted and untreated and CICAFLORA™ treated gatherings [40]. Biopsies treated with SPB1 LPs indicated completely re-epithelized wounds with amazing epidermal recovery. It has been, proposed that the free revolutionary rummaging properties of the LPs help to forestall inflammation and improve tissue arrangement, re- epithelization and separation of epidermis [41].

It has been proposed that the injury mending properties showed by those LPs researched might be because of their capacity to decrease oxidative pressure through the counteraction of receptive oxygen species creation.

5.3. Dermatological applications

The antibacterial additives utilized in most of individual consideration items are engineered and can cause skin bothering and hypersensitive responses by cooperation with keratin or collagen and elastin and empower the expulsion of lipids from the skin surface and influence the skin cells themselves. Biosurfactants are made out of lipid and proteins and are viable with the skin cell film. While most of biosurfactants related work is focused on biosurfactants that are created extra cellularly by miniature organic entities significantly less work has been completed on cell-bound biosurfactants a considerable lot of which are delivered by, for instance, probiotic Lactobacilli strains which have the additional favorable position of being nontoxic, biodegradable and harmless to the ecosystem [42-45].

The presentation of PEB was thought about against the glycolipids created by *Lactobacillus paracasei* (PAB). The PEB indicated antimicrobial action against *P. aeruginosa*, *Streptococcus agalactiae*, *S. aureus*, *E. coli*, *Streptococcus pyogenes* *C. albicans* and, which was tantamount with the outcomes from PAB. Significantly, separates arranged with phosphate-cradled saline (PBS) were more successful than phosphate support (PB) on account of *P. aeruginosa*, *S. aureus*, *E. coli* [46].

5.4. In Oral Care

In the common habitat, biosurfactants have been found to add to intrinsic oral consideration. Biosurfactants makers. In their investigation of the *S. mutans* viability of rhamnolipids got from non-pathogenic *Burholderia thailandensis*. The capability of et al. *B. subtilis* SPB1LP in toothpaste detailing and indicated that a L P-based item showed a significant antimicrobial action against *Enterobacter* SP and *S.typhimurium* . Past reports on the adequacy of SPB1 strain *B.subtilis* uncovered a wide range of activities including antimicrobial action towards miniature organic entities with MDR profiles, antifungal movement against et al. phytopathogenic parasites and antidiabetic and antilipidemic properties in alloxan-incited diabetic rodents [47].

5.5. Drug Delivery Systems (Including Vaccine)

The utilization of biosurfactants as medication conveyance specialists offers alluring applications, for example, uninvolved vaccination especially where drug treatment alternatives are restricted, the treatment of candidiasis is difficult because of the restricted accessibility of antifungal medications and their poison levels and serious results in people. These issues can be overwhelmed by fusing antifungal medications into different medication conveyance frameworks. Vesicular medication conveyance frameworks including liposomes and niosomes are believed to be especially significant for focused conveyance of medications and to limit unwanted results [48].

Liposomes remain as promising competitors with wide pertinence dependent on a medication conveyance approach including immunization, a kind of glycolipid biosurfactants that contains cationic liposomes has been appeared to advance quality transfection efficiency by five to multiple times with mam malian refined cells [49].

5.6. Antimicrobial and antifungal property

Given the ascent in anti-infection obstruction, the need to distinguish new antimicrobials and find a method for restoring current anti-microbials utilized in medication has gotten clear. There has been a worldwide invitation to battle as far as endeavours both broadly and universally to address the difficulty of anti-infection obstruction. Biosurfactants are obviously positioned to answer bring regarding their applications including: bactericidal, bacteriostatic, biofilm development restraint, biofilm disturbance, synergistic and adjuvant impacts with anti-microbials. Properties of biosurfactants incorporate hindrance of bacterial and contagious development. *Staphylococcus saprophyticus* antibacterial movement against *Klebsiella pneumonia*, *Escherichia coli*, *Vibrio cholera*, *Bacillus subtilis* *Staphylococandcus aureus*. Rhamnolipid has been accounted for to have biofilm problematic capacity against *Bacillus pumilus* [50].

The biosurfactant SUR can handle the development of in *Listeria mono cytogenes* food and some Gram-positive microorganisms like. LPs *B. pumulis*, *M.flavus* .can harm and enter lipid containing contrarily charged cell films. It has been recommended that a charge irregularity creates at the cell surface interface because of the polar component endeavoring to safeguard dissolvability. Deficiency of cell morphology prompting pore development in the lipid containing cell film of gram-negative microbes causing cell death (Fig. 2) [51].

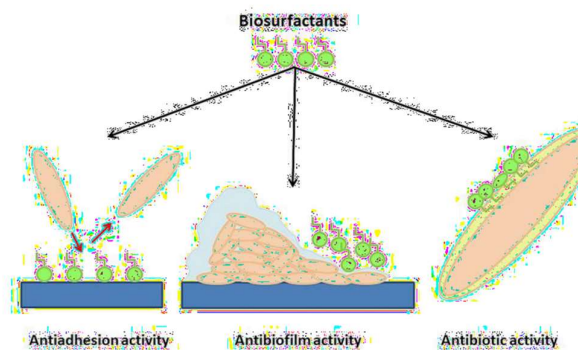


Fig. 2: Antiadhesive, antifilm and antibiotic activity of biosurfactants

6. ANALYSIS OF BIOSURFACTANS

There are various methods of analytical methods commonly used for various pharmaceutical formulations and bulk drugs. These methods are also used for the analysis of biosurfactants in the pure sample and in formulations. These methods include uv-spectroscopy, HPTLC, HPLC, gas chromatography, etc [52-95].

7. FUTURE PROSPECTIVE

Consequently, the potential applications regarding medical care therapeutics are substantially more encouraging given the worth added nature of such items and their probable benefit to human wellbeing. The expense benefits would have all the earmarks of being greater as far as the biomedical applications since creation is suitable on a limited scale. Scope of potential applications talked about here, all things considered, the intrinsic antimicrobial nature of numerous biosurfactants and the capacity of a portion of these to demonstration in cooperative energy as well as subordinates to current therapeutics with regards to the steadily expanding danger of anti-infection obstruction may demonstrate most beneficial (Table 3).

Table 3: Promising strategies of industrial biosurfactants production

SN	Strategy	Remarks (if any)
1	Use of growth enhance	Use of lactones enriches the production medium and enhance yield. Formation of a derivatized product makes recovery easier
2	Use of nanoparticles	Iron and manganese nanoparticles give enhance biosurfactant yield probably by replenishing the clinical metal ion requirements
3	Coproduction with other industrial product with another economical bioprocess	Bioprocessing for strain producing commercial enzyme along with biosurfactant would prove to be economical. Use of immobilized coculture for the same would also be equally cost effective
4	Use of immobilized producer organism	Fe nanoparticle enriched immobilizing medium, especially alginate, would provide easy separation and addition yield enhance with lesser by-product ducts. Activated charcoal acts as an enhancer as well as an immobilizing agent
5	Use of biofilm rector, pertraction and rotating discs bioreactor for recovery	Immobilization and aeration in rotating in rotating disc take care of foam production and subsequent loss of cells making recovery of product easier thereby reducing production cost.

8. CONCLUSION

From this discussion, it's evident that many medicinal plants exert an immunomodulatory effect in experimental models at a specific dose. Different types of in vivo and in vitro screening methods are employed in determining their pharmacological activity. Some medicinal plants may stimulate the system like *Ocimum sanctum*, *Tinospora cordifolia* and a few of them may suppress the immune responses example *Alternanthera tenella*. The review also reveals an update of the present immunomodulator plants and their pharmacological aspects. Thus, successful results are achieved by following an appropriate screening approach.

9. ACKNOWLEDGMENTS

We express our sincere thanks to Shri. Yogendraji Gode and Dr. Yogeshji Gode, IBSS's Dr. Rajendra Gode College of Pharmacy, Amravati and Dr. Rajendra Gode Institute of Pharmacy, Amravati.

10. DISCLOSURE OF CONFLICT OF INTEREST

The author declares no conflict of interest.

REFERENCES

1. Sing P, Patil Y. and. Rale V, biosurfactant production: emerging trends and production strategies. Journal of Applied Microbiology. 2018;126:2-13.
2. Naughton P.J., Marchant R., Naughton V. and Banat I.M. Microbial biosurfactant ; current trends and application in agriculture and biomedical industries, journal of applied microbiology 2019;127:12-28.
3. Nawazish Ali, Fenghuan Wang , Baocai Xu , Bushra Safdar , Asad Ullah , Muhammad Naveed Ce Wang and Muhammad Tayyab Rashid , Production and Application of Biosurfactant Produced by *Bacillus licheniformis* Ali5 in Enhanced Oil Recovery and Motor Oil Removal from Contaminated Sand , University of Ulster, Ulster , UK. Journal of Molecule 2019; 24:44-48.
4. Abdullahi Adekilekun Jimoh, Johnson Lin, Biosurfactant: A new frontier for greener technology and environmental sustainability, University of KwaZulu – Natal (Westville), Private Bag X54001.
5. McInerney M. J., Knapp R.M., Duncan Kathleen, Simpson D. R., Youssef N., Ravi N., Folmsbee M. J., Fincher T., Maudgalya S., Davis Jim, and Weiland Sandra, Development of an in situ biosurfactant production technology for enhanced oil recovery,
6. Banat, I.M., Franzetti, A., Gandolfi, I., Bestetti, G., Martinotti, M.G., Fracchia, L., Smyth, T.J. and Marchant, R. (2010) Microbial biosurfactants production, applications and future potential, 427-444. *Appl Microbiol Biotechnol* 87.
7. Banat, I.M., Satpute, S.K., Cameotra, S.S., Patil, R. and Nyayanit, N.V. (2014) Cost effective technologies and renewable substrates for biosurfactants' production. *Front Microbiol* 5, 1–18.
8. Cameotra, S.S., Makkar, R.S., Kaur, J. and Mehta, S.K. (2010) Synthesis of biosurfactants and their advantages to microorganisms and mankind. In ed. Sen, *Biosurfactants* R .pp. 261-280. New York: Springer.
9. Magdalena Pacwa - Płociniczak , Grażyna A. Płaza , Zofia Piotrowska-Seget and Swaranjit Singh Cameotra (2010-2011), Environmental Applications of Biosurfactants: Recent Advances, *International Journal of Molecular Sciences* ISSN 1422-0067, Department of Microbiology, Silesian University, Jagiellońska 28 street, 40-032 Katowice, Poland.
10. Amedea Perfumo, Ivo Rancich and Ibrahim M. Banat, Possibilities and Challenges for Biosurfactants Use in Petroleum Industry, *Biosurfactant*: edited by Ramkrishna Sen, Landes Bioscience and Springer Science+Business media 2010 ,(135-145).
11. Hall C, Tharakan P, Hallock J et al. Hydrocarbons and the evolution of human culture. *Nature* 2003; 426:318322.

12. Kanicky JR, Lopez-Montilla JC, Pandey S et al. Surface chemistry in the petroleum industry. In: Holmberg K,ed.
13. Handbook of Applied Surface and Colloid Chemistry. John Wiley and Sons Ltd, 2001:251-267.
14. Desai J, Banat IM. Microbial production of surfactants and their commercial potential. *Microbiol Mol Biol Rev* 1997; 61:47-64.
4. Banat IM, Makkar RS, Cameotra SS. Potential commercial applications of microbial surfactants. *Appl Microbiol Biotechnol* 2000; 53:495-508.
15. Jenneman GE, McInerney MJ, Knapp RM et al. A halotolerant, biosurfactant-producing bacillus species potentially useful for enhanced oil recovery. *Dev Ind Microbiol* 1983; 24:485-492.
16. Folmsbee M, Duncan K, Han SO et al. Re-identification of the halotolerant, biosurfactant-producing bacillus licheniformis strain JF-2 as *Bacillus mojavensis* JF-2. *Syst Appl Microbiol* 2006; 29:645-649.
17. McInerney MJ, Javaheri M, Nagle DP Jr. Properties of the biosurfactant produced by bacillus licheniformis strain JF-2. *J Ind Microbiol* 1990; 5:95-101.
18. Daoshan L, Shouliang L, Yi L et al. The effect of biosurfactant on the interfacial tension and adsorption loss of surfactant in ASP flooding. *Colloids and Surfaces A: Physicochem Eng Aspects* 2004; 244:53-60.
19. Gutnick D, Rosenberg E, Belsky I. et. al.-emulsans. US Patent 4,380,504 1983.
20. Magot M. Indigenous microbial communities in oil fields. In: Ollivier B, Magot M, eds. *Petroleum Microbiology*. Washington DC: ASM Press, 2005:21-33.
21. Javaheri M, Jenneman GE, McInerney MJ et al. Anaerobic production of a biosurfactant by bacillus licheniformis JF-2. *Appl Environ Microbiol* 1985; 50:698-700.
22. Rocha C, San-Blas F, San-Blas G et al. Biosurfactant production by two isolates of pseudomonas aeruginosa. *World Journal of Microbiol Biotechnol* 1992; 8:125-128.
23. Li Q, Kang C, Wang H et al. Application of microbial enhanced oil recovery technique to daqing oilfield. *Biochem Eng J* 2002; 11:197-199.
24. Ochsner UA, Koch AK, Fiechter A et al. Isolation and characterization of a regulatory gene affecting rhamnolipid biosurfactant synthesis in pseudomonas aeruginosa. *J Bacteriol* 1994; 176:2044-2054.
25. Youssef N, Simpson DR, Duncan KE et al. In situ biosurfactant production by bacillus strains injected into a limestone petroleum reservoir. *Appl Environ Microbiol* 2007; 73:1239-1247
26. Banat IM. Biosurfactant production and possible uses in microbial enhanced oil recovery and oil pollution remediation-a review. *Bioresour Technol* 1995; 51:112.
27. Bryant R., Potential use of microorganisms in petroleum recovery technology. *Proc Okla Acad Sci* 1987; 67:97104.
28. McInerney MJ, Nagle DP, Knapp RM. Microbially enhanced oil recovery: past, present and future. In: Ollivier B, Magot M, eds. *Petroleum Microbiology*. Washington DC: ASM Press, 2005:215-237.
29. Gutnick DL, Shabtai Y. Exopolysaccharide bioemulsifiers. In: Kosaric N, Cairns WL, Gray NCC, eds. *Biosurfactants and Biotechnology*. New York: Marcel Dekker Inc, 1987:211-246.
30. Hayes ME, Hrebenar KR, Murphy PL et al. Bioemulsifier-stabilized hydrocarbosols. US Patent 4,943,390, 1990.
31. Rocha CA, Gonzalez D, Iturralde ML et al. Production of oily emulsions mediated by microbial tenso-active agent.
32. US Patent 6,060,287, 2000.
33. Etoumi A. Microbial treatment of waxy crude oils for mitigation of wax precipitation. *J Pet Sci Eng* 2007; 55:111121.
34. Lazar I, Voicu A, Nicolescu C et al. The use of naturally occurring selectively isolated bacteria for inhibiting paraffin deposition. *J Pet Sci Eng* 1999; 22:161-169.

35. Gutnick D, Rosenberg E. Cleaning oil-contaminated vessels with -emulsans. US Patent 4,276,094, 1981.
36. Banat IM, Samarath N, Murad M et al. Biosurfactant production and use in oil tank clean-up. *World J Microbiol Biotechnol* 1991; 7:80-84.
37. Pesce L. A biotechnological method for the regeneration of hydrocarbons from dregs and muds, on the basis of biosurfactants. European Patent EP1427547, 2004.
38. Sahebazar, Z., Mowla, D. and Karimi, G. Enhancement of growth and Pseudomonas aeruginosa rhamnolipid production using iron-silica nanoparticles in low-cost medium. *J Nanostructures*. 2018, 8 ,1-10.
39. Obayori OS, Ilori, MO, Adebosoye, SA; Oyetibo, G.O.; Omotayo, A.E.; Amund, O.O. Degradation of hydrocarbons and biosurfactant production by Pseudomonas sp. strain LP1. *World J. Microbiol. Biotechnol.* 2009, 25, 1615–1623.
40. Reddy, M.S.; Naresh, B.; Leela, T.; Prashanthi, M.; Madhusudhan, N.C.; Dhanasri, G.; Devi, P. Biodegradation of phenanthrene with biosurfactant production by a new strain of Brevibacillus sp. *Bioresource Technol.* 2010, 101, 7980– 7983.
41. Kang, S.W.; Kim, Y.B.; Shin, J.D.; Kim, E.K. Enhanced biodegradation of hydrocarbons in soil by microbial biosurfactant, sophorolipid. *Appl. Biochem. Biotechnol.* 2010, 160, 780–790.
42. Das, K.; Mukherjee, A.K. Crude petroleum-oil biodegradation efficiency of Bacillus subtilis and Pseudomonas aeruginosa strains isolated from a petroleum-oil contaminated soil from North-East India. *Bioresource Technol.* 2007, 98, 1339– 1345.
43. Joseph, P.J.; Joseph, A. Microbial enhanced separation of oil from a petroleum refinery sludge. *J. Hazard. Mater.* 2009, 161, 522–525.
44. Barkay, T.; Navon-Venezia, S.; Ron, E.Z.; Rosenberg, E. Enhancement of solubilization and biodegradation of polyaromatic hydrocarbons by the bioemulsifier alasan. *Appl. Environ. Microbiol.* 1999, 65, 2697–2702.
45. Martínez-Checa, F.; Toledo, F.L.; Mabrouki, K.E.; Quesada, E.; Calvo, C. Characteristics of bioemulsifier V2-7 synthesized in culture media added of hydrocarbons: chemical composition, emulsifying activity and rheological properties. *Bioresour. Technol.* 2007, 98, 3130–3135.
46. Javaheri M, Jenneman GE, McInerney MJ et al. Anaerobic production of a biosurfactant by bacillus licheniformis JF-2. *Appl Environ Microbiol* 1985; 50:698-700.
47. Rocha C, San-Blas F, San-Blas G et al. Biosurfactant production by two isolates of pseudomonas aeruginosa. *World Journal of Microbiol Biotechnol* 1992; 8:125-128.
48. Li Q, Kang C, Wang H et al. Application of microbial enhanced oil recovery technique to daqing oilfield. *Biochem Eng J* 2002; 11:197-199.
49. Ochsner UA, Koch AK, Fiechter A et al. Isolation and characterization of a regulatory gene affecting rhamnolipid biosurfactant synthesis in pseudomonas aeruginosa. *J Bacteriol* 1994; 176:2044-2054.
50. Youssef N, Simpson DR, Duncan KE et al. In situ biosurfactant production by bacillus strains injected into a limestone petroleum reservoir. *Appl Environ Microbiol* 2007; 73:1239-1247
51. Banat IM. Biosurfactant production and possible uses in microbial enhanced oil recovery and oil pollution remediation-a review. *Bioresour Technol* 1995; 51:112.
52. Sabhadinde AF, et al. Novel RP-HPLC method for simultaneous analysis of chlorthalidone and telmisartan from combined dosage form. *Ijppr.Human.* 2020; 20(1):491-502.
53. Panchale WA, et al. RP-HPLC method for simultaneous determination of escitalopram oxalate and flupentixol HCl in tablet dosage form. *GSC Biological and Pharmaceutical Sciences.* 2021; 14(01):169-174.

54. Nimbokar SW, et al. Development and validation of RP-HPLC method for determination of zonisamide from tablet formulation. *World Journal of Pharmaceutical and Medical Research*. 2021;7(2):196-200.
55. Panchale WA, et al. RP-HPLC method for simultaneous determination of metformin hydrochloride and linagliptine in pharmaceutical dosage form. *World Journal of Pharmaceutical and Medical Research*. 2021;7(5):234- 238.
56. Manwar JV, et al. Development of newer RP-HPLC method for simultaneous estimation of cefixime and linezolid in bulk drugs and combined dosage form. *International Journal of Pharmacy and Life Sciences*. 2021;12(1):26-31.
57. Panchale WA, Gulhane CA, Manwar JV, Bakal RL. Simultaneous estimation of salbutamol sulphate and ambroxol HCl from their combined dosage form by UV-Vis spectroscopy using simultaneous equation method. *GSC Biological and Pharmaceutical Sciences*. 2020;13(03):127-134.
58. Bakal RL, et al. Spectrophotometric estimation of amitriptyline HCL and chlordiazepoxide in tablet dosage form. *International Journal of Chemical Sciences*. 2007; 5(1):360–364.
59. Panchale WA, Bakal RL. First-order derivative spectrophotometric estimation of gemifloxacin mesylate and ambroxol HCl in tablet dosage form. *GSC Biological and Pharmaceutical Sciences*. 2021; 14(2):029-036.
60. Gulhane CA, et al. Liquid chromatographic method for simultaneous estimation of thiocolchicoside and etoricoxib from tablet formulation. *Asian Journal of Pharmaceutical Analysis*. 2021;11(2): 118-122. DOI:10.52711/2231-5675.2021.00020.
61. Panchale WA, et al. Chromatographic analysis of famotidine, paracetamol and ibuprofen from tablet formulation. *Research Journal of Pharmacy and Technology*. 2019; 12:231-263.
62. Manwar JV, et al. Application of simultaneous equation method for the determination of azithromycin and cefixime trihydrate in tablet formulation. *Research Journal of Pharmacy and Technology*. 2017;10(1):108-112.
63. Manwar JV, et al. Response surface based optimization of system variables for liquid chromatographic analysis of candesartan cilexetil. *Journal of Taibah University for Science*. 2017; 11:159–172.
64. Manwar J, Mahadik K, Paradkar A, et al. Gas chromatography method for the determination of non-ethanol volatile compounds in herbal formulation. *International Journal of Analytical and Bioanalytical Chemistry*. 2013; 3(1):12-17.
65. Badukale NA, et al. Phytochemistry, pharmacology and botanical aspects of *Madhuca indica*: A review. *Journal of Pharmacognosy and Phytochemistry*. 2021; 10(2): 1280-1286.
66. Khadatkarn SN, et al. Preparations and evaluation of microcapsules of capsaicin. *International Journal of Chemical Sciences*. 2007; 5(5):2333-2341.
67. Panchale WA, et al. Concurrent analysis of ambroxol HCl and salbutamol sulphate from tablet formulation by RP-HPLC. *GSC Biological and Pharmaceutical Sciences*. 2020; 13(03):197-202.
68. Sahare AY, et al. *Hypericum perforatum*: A Medicinal plant. *Plant Archives*. 2007; 7(2):463-468.
69. Manmode R, et al. Effect of preparation method on antioxidant activity of ayurvedic formulation kumaryasava. *J Homeop Ayurv Med*. 2012;1:114. doi:10.4172/2167-1206.1000114
70. Padgilwar S, et al. Traditional uses, phytochemistry and pharmacology of *Oroxylum Indicum*: A Review. *International Journal of Pharmaceutical and Phytopharmacological Research*. 2014; 3 (6):483-486.
71. Manwar J, et al. Isolation, biochemical and genetic characterizations of alcohol-producing yeasts from the flowers of *Woodfordia fruticosa*. *J Young Pharm*. 2013;5(4):191-194.
72. Wadekar AB, et al. Morphology, phytochemistry and pharmacological aspects of *Carica papaya*, an review. *GSC Biological and Pharmaceutical Sciences*. 2020; 14(03):234-248.
73. Khadatkarn SN, et al. *In-vitro* anthelmintic activity of root of *Clitoria ternatea* linn. 2008; 4(13):148-150.

74. Sahare AY, et al. Antimicrobial activity of *Pseudarthria viscida* roots. Asian Journal of Microbiology Biotechnology & Environmental Sciences. 2008; 10(1):135-136.
75. Gudalwar BR, et al. Allium sativum, a potential phytopharmacological source of natural medicine for better health. GSC Advanced Research and Reviews. 2021; 06(03):220–232
76. Manwar JV, et al. Experimental design approach for chromatographic determination of ketorolac tromethamine from bulk drug and tablet formulation. Global Journal of Pharmacy & Pharmaceutical Sciences. 2017;3(2):38-47.
77. Malode GP, et al. Phytochemistry, pharmacology and botanical aspects of *Murraya Koenigii* in the search for molecules with bioactive potential - A review. GSC Advanced Research and Reviews. 2021; 06(03): 143–155.
78. Shubham Garibe, et al. Bioequivalence study of test formulations T1 and T2 Nadolol tablets USP with reference formulation in healthy adult, human subjects under fed conditions. Ijppr.Human. 2021; 20(2):20-28.
79. Chaudhari KD, et al. Comprehensive review on characterizations and application of gastro-retentive floating drug delivery system. GSC Advanced Research and Reviews. 2021; 07(01):035-044.
80. Chaudhari KD, et al. Floating drug delivery system: An update of preparation and classification of formulation. Ijppr.Human. 2021; 21 (1):207-220.
81. Manmode RS, et al. Stability indicating HPLC method for simultaneous determination of methocarbamol and nimesulide from tablet matrix. Der Chemica Sinica.2011;2(4):81-85.
82. Malode GP, et al. Formulation and evaluation of a novel floating in situ gel system for the treatment of peptic ulcer. World Journal of Pharmacy and Pharmaceutical Sciences 2021; 10(4):416-1433.
83. Jain CM, et al. Review on approaches for development and evaluation of extended-release tablets. Review on approaches for development and evaluation of extended-release tablets. World Journal of Pharmacy and Pharmaceutical Sciences 2021;10(4): 542-554.
84. Nimbawar MG, et al. A brief review on principle, preparation and properties of proniosomes: A vesicular drug delivery system. World J Pharm Sci. 2021; 9(5): 149-162.
85. Bagade SB, et al. Simultaneous high performance thin layer chromatographic estimation of methocarbamol and nimesulide in combined dose tablet. Journal of Pharmaceutical Research. 2006;5(4):137-140.
86. Vaidya VM, et al. Design and *in vitro* evaluation of mucoadhesive buccal tablets of terbutaline sulphate. Int J PharmTech Res. 2009; 1(3): 588-597.
87. Dhamankar AK, et al. The novel formulation design of O/of ketoprofen for improving transdermal absorption. Int J of Pharm Tech Res. 2009; 4(1Suppl): 1449-1457.
88. Manwar JV, et al. Diclofenac Sodium Loaded Nanosized Ethosomes: An Investigation on Z-Average, Polydispersity and Stability. J Pharm Res. 2017; 1(3): 000115.
89. Patil SS, et al. Ultrasound-Assisted Facile Synthesis of Nanostructured Hybrid Vesicle for the Nasal Delivery of Indomethacin: Response Surface Optimization, Microstructure, and Stability. AAPS PharmSciTech. 2019;20(3):97.
90. Parbat AY, et al. Ethnopharmacological review of traditional medicinal plants as immunomodulator. World Journal of Biology Pharmacy and Health Sciences, 2021, 06(02), 043–055.
91. Nimbawar MG, et al. Fabrication and evaluation of ritonavir proniosomal transdermal gel as a vesicular drug delivery system. Pharmacophore. 2016; 7(2): 82-95.
92. Pophalkar PB, et al. Development and evaluation of ondansetron medicated jelly. World Journal of Pharmaceutical Research. 2018; 7(19): 1252-1263.

93. Suroshe RS, et al. Development and characterization of osmotic drug delivery system of model drug. World Journal of Pharmaceutical Research. 2018; 7(18): 1158-1171.
94. Kadam CY, et al. Design and In vitro characterization of phase transition system using rivastigmine tartrate for nasal drug delivery system. World Journal of Pharmaceutical Research. 2018; 8(1): 815-829.
95. Gulhane CA, et al. UV- Visible Spectrophotometric estimation of azithromycin and cefixime from tablet formulation by area under curve method. World Journal of Pharmaceutical Sciences. 2021; 9(6): 163-168.