

PHYSICOCHEMICAL PROPERTIES AND TARGETING STRATEGIES FOR LIPOSOMES: A VERSATILE DRUG DELIVERY SYSTEM

Minakshee G. Nimbalwar <sup>1</sup>, Bhushan R. Gudalwar <sup>2</sup>, Anuja S. Motule <sup>1</sup>, Shivam A. Bartere <sup>2</sup>,  
Smita S. Sapate <sup>1</sup>, Jagdish V. Manwar <sup>2\*</sup> and Ravindra L. Bakal <sup>1</sup>

<sup>1</sup> IBSS's Dr. Rajendra Gode Institute of Pharmacy, Mardi road, Amravati-444 602, MS, India.

<sup>2</sup> IBSS's Dr. Rajendra Gode College of Pharmacy, Mardi Road, Amravati-444 602, MS, India.

\*Corresponding Author: Email: [jvmanwar@gmail.com](mailto:jvmanwar@gmail.com)

Received: 26 August 2021 / Revised: 23 November 2021 / Accepted: 22 December 2021 / Available online: 31 December 2021

## ABSTRACT

*Liposomes are phospholipid-based bilayer vesicular systems. It can encapsulate both hydrophilic and hydrophobic drugs. Its properties like biocompatibility and site-specific action are the two most attractive features of the systems. Their architecture has been satisfactorily accepting a great technical advance such as remote drug loading, combining passive, active drug targeting concepts, etc. It can be used in diverse areas like carrier for anticancer, bio-active molecules, diagnostics, and therapeutic agents. In present review, we discuss in detailed about structure of liposomes and liposomes as effective targeting delivery system.*

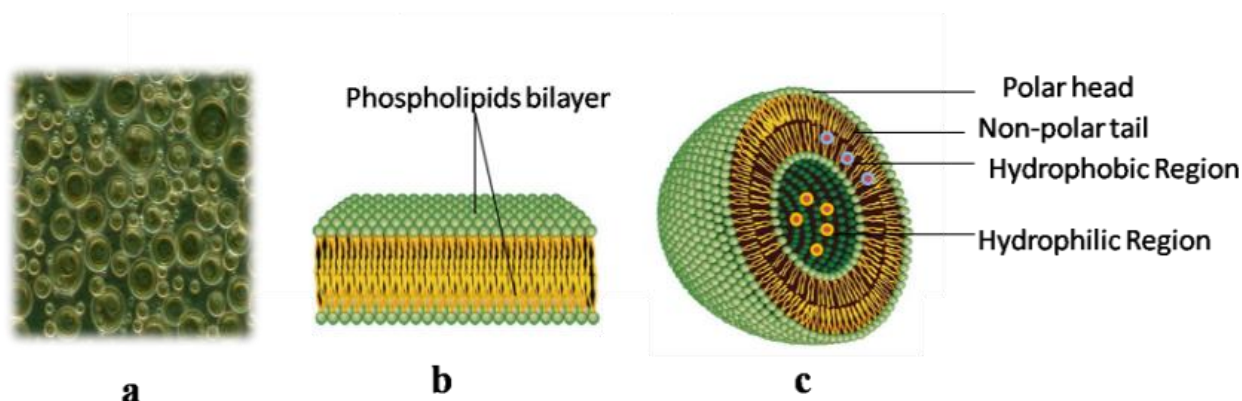
**Keywords** – Liposomes; Vesicle; Physicochemistry; Targeting site.

## 1. INTRODUCTION

Development of suitable drug delivery system has become a challenging task to deliver particular type of drug to the targeted site. There are number of drug delivery systems so far developed for various drugs. These systems include ethosomes, liposomes, tablets, multilayered tablets, emulsion, suspension, etc. New drug delivery Liposomes are phospholipid-based bilayer vesicular systems. They are discovered by Alec Bangham in 1964. It was found that phospholipids in aqueous systems can form closed bilayered spherical vesicular structures (**Fig.1**) [1-27]. They have emerged as one of the most promising tools for drug targeting in medical fields. Mostly it is composed of natural substances like cholesterol, phosphatidylglycerol, dicetylphosphate, distearoylphosphatidylcholine, dipalmitoylphosphatidylcholine, etc. It is nontoxic, biodegradable, non-immunogenic and biocompatible. The size varies from a few nanometers to several micrometers [28-29].

In medical use, the range of liposomes applied in between 50-450 nm. The properties of liposomes are mostly dependent on lipid composition, surface charge, size, and the method of preparation. The rigidity of bilayer and the charge are depending on the selection of the bilayer components [30-32]. Conventional liposomes tend to form aggregate with each other resulting in immature release of liposomal contents over time [33-34]. To overcome such problems associated with liposomes, the surface-

modification strategies were adopted by coating the surface of liposomes with inert, biocompatible hydrophilic polymers, such as polyethylene glycol. These polymers confer steric stabilization to the liposomal surface via formation of a protective layer over the liposome surface and slows down liposome recognition by opsonins, and therefore, subsequent clearance of liposomes [35-45].



**Fig. 1: Liposomes (a) Microscopic, (b) Phospholipid bilayer, and (c) Spherical Vesicle.**

## **2. PHYSICOCHEMISTRY OF SYSTEM**

Liposomes are the drug carrier for delivering both the hydrophilic and the lipophilic drugs. The strength of liposomes as a carrier system for drugs depend on the number of physicochemical properties of the liposomal membranes, on the nature of their components, their size, surface charge, and the lipid organization [46]. As liposomes composed of phospholipids, dispersed in aqueous phase, due to their amphiphatic nature they have a strong tendency to form membranes by polar head and nonpolar tail [47]. Polar heads prefer to interact with the aqueous environment and long non-polar aliphatic chains promote interaction with one another (**Fig.1**) [48].

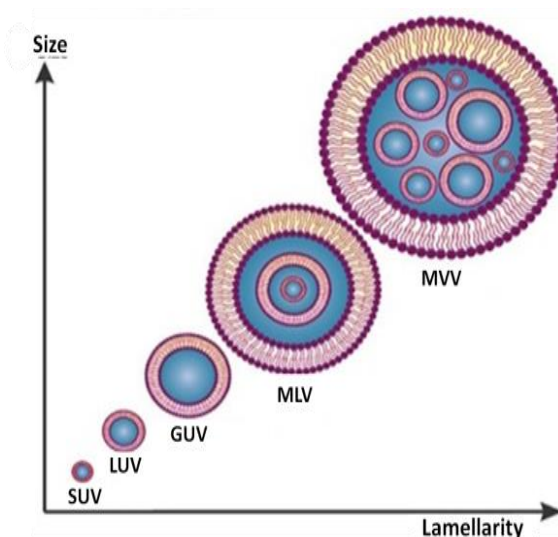
Hydrophobic interactions are behind the formation of these lipid bilayers and the forces like Van der Waals keep the long hydrocarbon tails together, thus it helps for strengthening of this architecture. Stabilization of this vesicle is due to hydrogen bonds and polar interactions between the water molecules of the aqueous environment and the polar heads of an organization. Whole structural organization of lipids depends on their nature, concentration, temperature, and geometric form. If ions or other drug molecules are present during the formulation process of liposomes, they can be encapsulated inside these membranes [49-50].

The addition of cholesterol in the liposomal formulation reduces their permeability and increases its stability by interacting with the core of the membrane, because the presence of cholesterol induces a dense packing of phospholipids and inhibits their transfer to high-density lipoprotein and low-density lipoprotein [51]. Also, the use of phosphatidyl choline with saturated fatty acyl chains and materials that stretch the transition temperature beyond 37°C important for better stabilization [52-54].

## **3. CLASSIFICATION**

Liposomes can be classified on the basis of the method of preparation like reverse-phase evaporation vesicles or vesicle extruded technique, size small, intermediate, or large, and lamellarity uni-, oligo-, and multilamellar vesicles (**Fig. 2**). The formation of

unilamellar vesicles (ULVs) composed of one lipid bilayer with size 50–250 nm. Ideally it is important for the encapsulation of hydrophilic drugs [55].



**Fig. 2: Liposomes according to size and lamellarity.**

The multilamellar vesicles (MLVs) depend on the synthesis methods and post formation processing used for their preparation. It is composed of two or more concentric lipid bilayers organized by packing of an onion, having size 1-5µm preferentially entrap lipid-soluble drugs [56]. The multi vesicular vesicles (MVV) composed of multiple vesicles entrapped under the lipid bilayer. According to the lamellar arrangement MLVs are formed more easily at larger hydrodynamic diameters. It has greater entrapped volume than ULVs. [57] **(Table 1)**.

**Table 1: Types of liposomes with their size and lipid layers [58].**

Vesicle Type	Number of lipid bilayers	Size of vesicles
Small Unilamellar Vesicles (SUV)	1	50-100 nm
Large Unilamellar Vesicles (LUV)	1	All size ranges
Giant Unilamellar Vesicles (GUV)	1	More than 1 micrometer
Multilamellar Vesicles (MLV)	5-25	More than 0.5 micrometer
Multi Vesicular Vesicles (MLV)	Multi compartmental str.	More than 0.5micrometer

#### **4. FORMATION OF TARGETED LONG-CIRCULATING LIPOSOMES**

Targeted long circulating liposome is a critical step to achieve development of liposomes. It must survive in the systemic circulation for longer time to reach and bind to the targeted site. Strategies to increase the circulation time of liposomes, that is reduction in liposomal size or the inclusion of cholesterol and high phase transition lipids provided a modest decrease in the

clearance rate of liposomes. More success came from the strategy of including a hydrophilic molecule at the liposomal surface, e.g., phosphatidylinositol or lipid derivatives of polymers like PEG, poly(acrylamide), poly(vinylpyrrolidone), etc. [59].

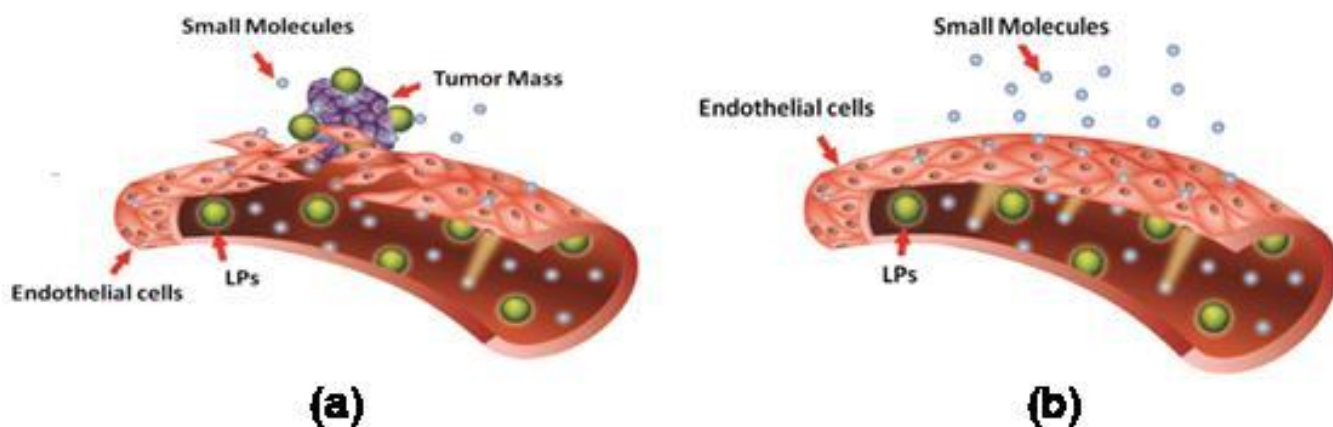
Coupling strategies of molecule is via cross-linking molecules of a ligand is coupled, often through a spacer molecule, to a hydrophobic molecule. The stable insertion of the conjugate into the lipid bilayer of a liposomes is important. Generally, the carrier of choice has been phosphatidylethanolamine (PE) because of the reactive amine in its head group and the availability of various acyl chain lengths of different degrees of unsaturation. PEG- at concentrations of approximately 5% or more in the bilayer, probably sterically interfere with the accessibility of the antibody to the liposomal surface. PEG could be incorporated into liposomes after ligand coupling occurs, to overcome the interference of PEG in the conjugation of antibodies onto the liposomal surface [60-62].

## 5. LIPOSOME TARGETING STRATEGIES

### A. Passive targeting

Drugs associated with a carrier, exhibit two pharmacokinetic processes related to the liposomal system and the drug itself. When the drug is released slowly, the kinetics will be determined essentially by that of the liposomes. Accordingly, drugs with short biological half-lives, such as retinoic acid and Curcumin owing to rapid clearance can be encapsulated into a liposome which protects the drug providing minimal to no leakage [63]. The macrophagial uptake requires binding of specific serum proteins to the drug-particulate system for identification and consequent elimination, a phenomenon commonly known as opsonization [64].

The most widely used polymers to formulate stealth liposomes are PEG-based conjugates, it has unique characteristics of this polymer including non-ionic nature, high solubility in both aqueous and organic media, excellent biocompatibility, lack of toxicity, low immunogenicity and antigenicity, good excretion kinetics, as well as ease of synthesis with a broad range of molecular weight and a low polydispersity index is  $< 1.1$  and a variety end-group functionalities [65-66]. Although other surface modifying polymers are much less common, a recent patent claimed that liposomes stabilized by poly 2-methacryloyloxyethyl phosphorylcholine can enhance retention *in vivo* to a greater extent than do PEGylated liposomes [67-69].



**Fig. 3: Targeting of LPs on tumor (a) and normal (b) vessels by enhanced permeability and retention effect.**

## **B. Ligand-mediated targeting**

Ligand or receptor-mediated targeting is based on the surface charges of the particle with ligands that bind specifically to receptors. Ligand-receptor binding initiates the uptake of the liposomes by the targeted cell. Targeting ligands such as proteins, peptides, antibodies, antibody fragments, vitamins, carbohydrates and glycoproteins have shown the ability to bind selectively target cells such as tumor cells. For e.g., ginsenosides which are substrates of glucose-related transporters overexpressed in certain tumors and have the ability to cross the blood-brain barrier (BBB) via glucose transporter 1-mediated transport can therefore function as targeting ligands [70-72].

When the objective is imaging a specific target, it is crucial for the imaging agent to accumulate at the desired site in sufficient concentration. A possible strategy to enhance imaging outcomes is the use of ligand-mediated targeting liposomes, carrying a suitable imaging agent, which exhibit the ability to bind specifically to the receptor of interest [73-74]. The BBB, which is formed by endothelial capillary endothelium and separates the vascular system from the brain, is a strict permeability barrier that efficiently restricts penetration of therapeutic compounds to the brain. A fusion protein containing a six amino acid peptide with the ability to target the BBB has been incorporated in a liposomal formulation, comprising CHOL: phosphatidyl phosphoric acid:PC (30-50:5-20:40-60), in which the targeting protein is anchored and conjugated to 1,2-dioleoyl-sn-glycero-3-succinate within the LP bilayer [75-76].

## **C. Combined targeting**

Ligand-mediated targeting would require extended residence time in blood of the liposomal system for an effective access and accumulation at the site of action. Therefore, it is very common to engineer therapeutic liposomes to exhibit combined passive and ligand-mediated targeting capabilities (passive + ligand-mediated). Combined targeting is attained by functionalizing the surface of liposomes with both a targeting ligand and a hydrophilic polymer, mostly PEG chains, in which the ligand is usually coupled to the distal end of the polymeric chain. The PEG linkers may enhance ligand's binding affinity for the receptors and the flexible PEG spacers are known to enhance ligand-receptor binding. However, it was shown that the distal conjugation of the trigalactosyl on the PEG chain reduced the uptake of the particle by hepatocytes by comprising with the non-PEGylated ligand [77-79].

## **D. Stimuli-responsive liposomes**

Stimuli-responsive LPs are intelligent systems that have the ability to deliver and release their cargo in a site-specific manner. The composition of these liposomal systems can be specifically designed to facilitate triggered or controlled release in response to an endogenous stimulus like pH, redox potential, or specific enzymatic activity or an applied external stimulus these are heat, light and ultrasound (US) waves. Triggered drug release is based on the concept of membrane destabilization induced by local defects within bilayer membranes to promote payloads release [80].

## **6. METHODS OF ASSAY OF LIPOSOMES**

There are many analytical tools that are used for the analysis of various pharmaceutical drugs, formulations, including liposomes. These methods include UV-Vis spectrophotometry, gas chromatography, HPLC, HPTLC, etc. [81- 115].

## **7. CONCLUSION**

Liposomal drug delivery systems can be used for delivery of both hydrophilic and lipophilic drugs. Thus, it concludes that it could be effectively used in delivery of drugs for severe diseases.

## **8. DISCLOSURE OF CONFLICT OF INTEREST**

The author declares no conflict of interest.

## **REFERENCES**

1. Nimbawar MG, Panchale WA, Nimbokar SW, Gudalwar BR, ManwarJV, Bakal RL. A brief review on principle, preparation and properties of proniosomes: A provesicular drug delivery system. *World J Pharm Sci.* 2021; 9(5): 149-162.
2. Chaudhari KD, et al. Comprehensive review on characterizations and application of gastro-retentive floating drug delivery system. *GSC Advanced Research and Reviews.* 2021; 07(01): 035-044.
3. Chaudhari KD, et al. Floating drug delivery system: An update of preparation and classification of formulation. *Ijppr.Human.* 2021; 21(1): 207-220.
4. Gudalwar BR, et al. Allium sativum, a potential phytopharmacological source of natural medicine for better health. *GSC Advanced Research and Reviews.* 2021; 06(03): 220–232.
5. Dhamankar AK, et al. The novel formulation design of O/of ketoprofen for improving transdermal absorption. *Int J of Pharm Tech Res.* 2009; 4(1Suppl): 1449-1457.
6. Dongare PN, Motule AS, et al. Recent development in novel drug delivery systems for delivery of herbal drugs: An updates. *GSC Advanced Research and Reviews.* 2021; 8(08):008-018.
7. Upadhye KP, Senpal D, Nimbawar M, Dixit G. Formulation and evaluation of fish oil-based rizatriptan microemulsion for intranasal migraine treatment. *International Journal of Pharmaceutical Sciences and Nanotechnology.* 2015;8(30): 2972-2978.
8. Gulhane CA, Motule AS, et al. An overview on nail drug delivery system: A Promising application for various diseases. *European Journal of Biomedical and Pharmaceutical Sciences.* 2021;8(2):104-110.
9. Jain CM, et al. Review on approaches for development and evaluation of extended-release tablets. Review on approaches for development and evaluation of extended-release tablets. *World Journal f Pharmacy and Pharmaceutical Sciences.* 2021;10(4): 542-554.
10. Kadam CY, et al. Design and *In vitro* characterization of phase transition system using rivastigmine tartrate for nasal drug delivery system. *World Journal of Pharmaceutical Research.* 2018; 8(1): 815-829.
11. Khadatkar SN, et al. *In-vitro* anthelmintic activity of root of Clitoria ternatea linn. 2008; 4(13): 148-150.

12. Khadatkar SN, et al. Preparations and evaluation of microcapsules of capsaicin. *International Journal of Chemical Sciences*. 2007; 5(5): 2333-2341.
13. Malode GP, et al. Formulation and evaluation of a novel floating in situ gel system for the treatment of peptic ulcer. *World Journal of Pharmacy and Pharmaceutical Sciences*. 2021; 10(4): 416-1433.
14. Manmode R, et al. Effect of preparation method on antioxidant activity of ayurvedic formulation kumaryasava. *J Homeop Ayurv Med*. 2012; 1: 114.
15. Manwar J, et al. Isolation, biochemical and genetic characterizations of alcohol-producing yeasts from the flowers of *Woodfordia fruticosa*. *J Young Pharm*. 2013; 5(4): 191-194.
16. Manwar JV, et al. Diclofenac Sodium Loaded Nanosized Ethosomes: An Investigation on Z-Average, Polydispersity and Stability. *J Pharm Res*. 2017; 1(3): 000115.
17. Motule, AS, et al. Development and physicochemical evaluation of bilayered transdermal patches of ondansetron hydrochloride. *Journal of Innovations in Pharmaceutical and Biological Sciences*. 2021; 8(3): 17-23.
18. Nimbawar MG, et al. A brief review on principle, preparation and properties of proniosomes: A vesicular drug delivery system. *World J Pharm Sci*. 2021; 9(5): 149-162.
19. Nimbawar MG, et al. Fabrication and evaluation of ritonavir proniosomal transdermal gel as a vesicular drug delivery system. *Pharmacophore*. 2016; 7(2): 82-95.
20. Nimbawar MG, Gudalwar BR, Panchale WA, WadekarAB, Manwar JV, Bakal RL. An overview of characterizations and applications of proniosomal drug delivery system. *GSC Advanced Research and Reviews*. 2021; 07(02): 025–034.
21. Parbat AY, et al. Ethnopharmacological review of traditional medicinal plants as immunomodulator. *World Journal of Biology Pharmacy and Health Sciences*. 2021; 06(02): 043–055.
22. Patil SS, et al. Ultrasound-Assisted Facile Synthesis of Nanostructured Hybrid Vesicle for the Nasal Delivery of Indomethacin: Response Surface Optimization, Microstructure, and Stability. *AAPS PharmSciTech*. 2019; 20(3): 97.
23. Pophalkar PB, et al. Development and evaluation of ondansetron medicated jelly. *World Journal of Pharmaceutical Research*. 2018; 7(19): 1252-1263.
24. Sahare AY, et al. Antimicrobial activity of *Pseudarthria viscida* roots. *Asian Journal of Microbiology Biotechnology & Environmental Sciences*. 2008; 10(1): 135-136.
25. Shubham Garibe, et al. Bioequivalence study of test formulations T1 and T2 Nadolol tablets USP with reference formulation in healthy adult, human subjects under fed conditions. *Ijppr.Human*. 2021; 20(2): 20-28.
26. Suroshe RS, et al. Development and characterization of osmotic drug delivery system of model drug. *World Journal of Pharmaceutical Research*. 2018; 7(18): 1158-1171.

27. Vaidya VM, et al. Design and in vitro evaluation of mucoadhesive buccal tablets of terbutaline sulphate. *Int J PharmTech Res.* 2009; 1(3): 588-597.
28. Giuseppina B, Agnese M. Liposomes as nanomedical devices. *International Journal of Nanomedicine* 2015;10: 975-999
29. Bangham AD, Hill MW, Miller NG. Preparation and use of liposomes as models of biological membranes. In: Korn ED, editor. *Methods in Membrane Biology.* Vol 1. New York: Plenum; 1974:1–68.
30. Bangham AD, Horne RW. Negative staining of phospholipids and their structural modification by surface-active agents as observed in the electron microscope. *J Mol Biol.* 1964; 8:660–668.
31. Pattni BS, Chupin VV, Torchilin VP. New developments in liposomal drug delivery. *Chem Rev.* 2015; 115:10938–66.
32. Maja, L., Zeljko, K., & Mateja, P. Sustainable technologies for liposome preparation. *J. of Supercritical Fluids*2020; 165:104984.
33. Zylberberg C, Matosevic S. Pharmaceutical liposomal drug delivery: a review of new delivery systems and a look at the regulatory landscape. *Drug Deliv.* 2016;23:3319–29.
34. Kim JS. Liposomal drug delivery system. *J Pharmaceut Invest.* 2016; 46:387–92.
35. Sercombe L, Veerati T, Moheimani F, et al. Advances and challenges of liposome assisted drug delivery. *Front Pharmacol.* 2015;6:286.
36. Haluska CK, Riske KA, Marchi-Artzner V, Lehn JM, Lipowsky R, Dimova R. Time scales of membrane fusion revealed by direct imaging of vesicle fusion with high temporal resolution. *Proc. Natl. Acad. Sci. U.S.A.*, 103, 15841–15846 (2006).
37. Bulbake U, Doppalapudi S, Kommineni N, et al. Liposomal formulations in clinical use: an updated review. *Pharmaceutics.* 2017;9(2).
38. Ni, Y., Kong, L., Li, X., Xiao, H., Wu, Y., Liang, X., Yang, J. Multifunctional osteole liposomes and brain targeting functionality with potential applications in a mouse model of Alzheimer’s disease. *Journal of Liposome Research*, 2020;1–32.
39. Li M, Du C, Guo N, et al. Composition design and medical application of liposomes. *Eur J Med Chem.* 2019;164:640–53.
40. Nimbawar MG, Panchale WA, Nimbokar SW, Gudalwar BR, ManwarJV, Bakal RL. A brief review on principle, preparation and properties of proniosomes: A provesicular drug delivery system. *World J Pharm Sci.* 2021; 9(5): 149-162.
41. Gabizon A, Shmeeda H, Barenholz Y. Pharmacokinetics of pegylated liposomal doxorubicin: review of animal and human studies. *Clin. Pharmacokinet.*, 2003; 42: 419–436
42. Tuffin G, Waelti E, Huwyler J, Hammer C, Marti HP. Immunoliposome targeting to mesangial cells: a promising strategy for specific drug delivery to the kidney. *J. Am. Soc. Nephrol.*, 2005;16: 3295–3305.
43. Paszko E, Senge MO. Immunoliposomes. *Curr. Med. Chem.*, 2012; 19:5239–5277.



44. Barenholz Y. Doxil(R)—the first FDA-approved nano-drug: lessons learned. *J. Control. Release*, 2012; 160:117–134 .
45. Abu Lila AS, Kizuki S, Doi Y, Suzuki T, Ishida T, Kiwada H. Oxaliplatin encapsulated in PEG-coated cationic liposomes induces significant tumor growth suppression via a dual-targeting approach in a murine solid tumor model. *J. Control. Release*, 2009; 137: 8–14.
46. Polisky BA, Adami RC, Templin MV, et al.; Marina Biotech Inc. Processes and compositions for liposomal and efficient delivery of gene silencing therapeutics. EP2902013, 2015.
47. Euliss LE, DuPont JA, Gratton S, DeSimone J. Imparting size, shape, and composition control of materials for nanomedicine. *Chem Soc Rev*. 2006; 35:1095–1104.
48. Papahadjopoulos D, Kimelberg HK. Phospholipid vesicles (liposomes) as models for biological membranes: their properties and interactions with cholesterol and proteins. In: *Progress in Surface Science*. Vol. Oxford: Pergamon; 1973:141–149.
49. Chu CJ, Dijkstra J, Lai MZ, Hong K, Szoka FC. . Efficiency of cytoplasmic delivery by pH-sensitive liposomes to cells in culture. *Pharm Res*. 1990;7:824–834.
50. Hosta-Rigau L, Zhang Y, Teo BM, Postma A, Städler B. Cholesterol – a biological compound as a building block in bionanotechnology. *Nanoscale*. 2013; 5:89–109.
51. Bitounis D, Fanciullino R, Iliadis A, Ciccolini J. Optimizing druggability through liposomal formulations: new approaches to an old concept. *ISRN Pharm*. 2012; 2012:738432.
52. Milla P, Dosio F, Cattel L. PEGylation of proteins and liposomes: a powerful and flexible strategy to improve the drug delivery. *Curr Drug Metab*. 2012; 13:105–109.
53. Gabizon A, Papahadjopoulos D. Liposome formulations with prolonged circulation time in blood and enhanced uptake by tumors. *Proc Natl Acad Sci U S A*. 1988; 85:6949–6953.
54. Immordino ML, Dosio F, Cattel L. Stealth liposomes: review of the basic science, rationale, and clinical applications, existing and potential. *Int J Nanomedicine*. 2006; 1:297–315.
55. Betageri GV, Parsons DL. Drug encapsulation and release from multilamellar and unilamellar liposomes. *Int J Pharm*. 1992; 81:235–241.
56. Niven RW, Speer M, Schreier H. Nebulization of liposomes. II. The effects of size and modeling of solute release profiles. *Pharm Res*. 1991; 8:217–221.
57. Immordino ML, Dosio F, Cattel L. Stealth liposomes: review of the basic science, rationale, and clinical applications, existing and potential. *Int J Nanomedicine*. 2006; 1:297.
58. Allen TM. Long-circulating (Stealth) liposomes: therapeutic applications. In: Puisieux F, Couvreur P, Delattre J, Devissaguet JP, editors. *Liposomes: New Systems and New Trends in Their Applications*. Paris: Editions de Santé; 1995:125–155.

59. Mohan A, Narayanan S, Sethuraman S, Krishnan UM. Novel resveratrol and 5-fluorouracil coencapsulated in pegylated nanoliposomes improve chemotherapeutic efficacy of combination against head and neck squamous cell carcinoma. *Biomed Res Int.* 2014; 2014: 424239.
60. Storm G, Roerdink FH, Steerenberg PA, de Jong WH, Crommelin DJ. Influence of lipid composition on the antitumor activity exerted by doxorubicin-containing liposomes in a rat solid tumor model. *Cancer Res.* 1987; 47:3366–3372.
61. Cullis PR, Hope MJ. The bilayer stabilizing role of sphingomyelin in the presence of cholesterol: a <sup>31</sup>P NMR study. *Biochim Biophys Acta.* 1980; 597:533–542.
62. Defrise-Quertain F, Chatelain P, Delmelle M, et al. Model studies for drug entrapment and liposome stability. In: Gregoriadis G, editor. *Liposome Technology.* Boca Raton (FL): CRC; 1984:1–17.
63. Deodhar S, Dash AK. Long circulating liposomes: challenges and opportunities. *Ther Deliv.* 2018; 9:857–72.
64. Lee SG, Gangangari K, Kalidindi TM, et al. Copper-64 labeled liposomes for imaging bone marrow. *NuclMed Biol.* 2016; 43:781–7.
65. Tan S, Zou C, Zhang W, et al. Recent developments in d- $\alpha$ -tocopheryl polyethylene glycol-succinate-based nanomedicine for cancer therapy. *Drug Deliv.* 2017; 24:1831–42.
66. Nag OK, Awasthi V. Surface engineering of liposomes for stealth behavior. *Pharmaceutics.* 2013; 5:542–569.
67. Cheng Z, Al Zaki A, Hui JZ, et al. Multifunctional nanoparticles: cost versus benefit of adding targeting and imaging capabilities. *Science.* 2012; 338:903–10.
68. Bertrand N, Wu J, Xu X, et al. Cancer nanotechnology: the impact of passive and active targeting in the era of modern cancer biology. *Adv Drug Deliv Rev.* 2014; 66:2–25.
69. Natfji AA, Ravishankar D, Osborn HMI, et al. Parameters affecting the enhanced permeability and retention effect: the need for patient selection. *J Pharm Sci.* 2017; 106:3179–3187.
70. Johnsen KB, Burkhart A, Melander F, et al. Targeting transferrin receptors at the blood-brain barrier improves the uptake of immunoliposomes and subsequent cargo transport into the brain parenchyma. *Sci Rep.* 2017; 7:10396.
71. Akopian V, Minomi K, Niitsu Y, et al.; Nitto Denko Corp. Retinoid-liposomes for enhancing modulation of HSP47 expression. EP3075855; 2014.
72. Toporkiewicz P, Meissner J, Kuliczowski K, et al.; Wrocławskie Centrum Badań EIT+ SP. Z O.O. Lipid composition used for construction of liposomal genetic drug carrier targeted with antibodies, and use thereof. WO2015160271: 2015 October 22.
73. Niyikiza C, Varghese J; L.E.A.F. Holdings Group Llc. Liposome encapsulated affinity drug. WO2016025882; 2016 February 18.
74. Pisano C, Cecere SC, Di Napoli M, et al. Clinical trials with pegylated liposomal doxorubicin in the treatment of ovarian cancer. *J Drug Deliv.* 2013; 2013:898146.

75. Gordon AN, Granai CO, Rose PG, et al. Phase II study of liposomal doxorubicin in platinum- and paclitaxel-refractory epithelial ovarian cancer. *J Clin Oncol.* 2000; 18:3093–3100.
76. Kusumoto K, Harashima H, Akita H, et al.; Hokkaido University, Taiho Pharmaceutical Co Ltd. Vector for pulmonary delivery, inducing agent, and uses. EP2687204; 2014.
77. Santiwarangkool S, Akita H, Nakatani T, et al. PEGylation of the GALA peptide enhances the lung-targeting activity of nanocarriers that contain encapsulated siRNA. *J Pharm Sci.* 2017;106:2420–7
78. Wu M, Huang T, Wang J, et al. Antitumor cancer effect of ergosterol and cisplatin-loaded liposomes modified with cyclic arginine-glycine-aspartic acid and octa-arginine peptides. *Medicine (Baltimore).* 2018;97:11916.
79. Lee Y, Thompson DH. Stimuli-responsive liposomes for drug delivery. *Wiley Interdiscip Rev Nanomed Nanobiotechnol.* 2017;9(5).
80. Frisch B, Carriere M, Largeau C, et al. A new triantennary galactose-targeted PEGylated gene carrier, characterization of its complex with DNA, and transfection of hepatoma cells. *Bioconjug Chem.* 2004;15:754–64.
81. Sabhadinde AF, et al. Novel RP-HPLC method for simultaneous analysis of chlorthalidone and telmisartan from combined dosage form. *Ijppr.Human.* 2020; 20(1): 491-502.
82. Panchale WA, et al. RP-HPLC method for simultaneous determination of escitalopram oxalate and flupentixol HCl in tablet dosage form. *GSC Biological and Pharmaceutical Sciences.* 2021; 14(01): 169-174.
83. Nimbokar SW, et al. Development and validation of RP-HPLC method for determination of zonisamide from tablet formulation. *World Journal of Pharmaceutical and Medical Research.* 2021; 7(2): 196-200.
84. Panchale WA, et al. RP-HPLC method for simultaneous determination of metformin hydrochloride and linagliptine in pharmaceutical dosage form. *World Journal of Pharmaceutical and Medical Research.* 2021; 7(5): 234- 238.
85. Manwar JV, et al. Development of newer RP-HPLC method for simultaneous estimation of cefixime and linezolid in bulk drugs and combined dosage form. *International Journal of Pharmacy and Life Sciences.* 2021; 12(1): 26-31.
86. Panchale WA, Gulhane CA, Manwar JV, Bakal RL. Simultaneous estimation of salbutamol sulphate and ambroxol HCl from their combined dosage form by UV-Vis spectroscopy using simultaneous equation method. *GSC Biological and Pharmaceutical Sciences.* 2020; 13(03): 127-134.
87. Bakal RL, et al. Spectrophotometric estimation of amitriptyline HCl and chlordiazepoxide in tablet dosage form. *International Journal of Chemical Sciences.* 2007; 5(1): 360–364.
88. Panchale WA, Bakal RL. First-order derivative spectrophotometric estimation of gemifloxacin mesylate and ambroxol HCl in tablet dosage form. *GSC Biological and Pharmaceutical Sciences.* 2021; 14(2): 029-036.
89. Gulhane CA, et al. Liquid chromatographic method for simultaneous estimation of thiocolchicoside and etoricoxib from tablet formulation. *Asian Journal of Pharmaceutical Analysis.* 2021; 11(2): 118-122.

90. Panchale WA, et al. Chromatographic analysis of famotidine, paracetamol and ibuprofen from tablet formulation. *Research Journal of Pharmacy and Technology*. 2019; 12: 231-263.
91. Manwar JV, et al. Application of simultaneous equation method for the determination of azithromycin and cefixime trihydrate in tablet formulation. *Research Journal of Pharmacy and Technology*. 2017; 10(1): 108-112.
92. Manwar JV, et al. Response surface based optimization of system variables for liquid chromatographic analysis of candesartan cilexetil. *Journal of Taibah University for Science*. 2017; 11: 159–172.
93. Manwar J, Mahadik K, Paradkar A, et al. Gas chromatography method for the determination of non-ethanol volatile compounds in herbal formulation. *International Journal of Analytical and Bioanalytical Chemistry*. 2013; 3(1): 12-17.
94. Badukale NA, et al. Phytochemistry, pharmacology and botanical aspects of *Madhuca indica*: A review. *Journal of Pharmacognosy and Phytochemistry*. 2021; 10(2): 1280-1286.
95. Panchale WA, et al. Concurrent analysis of ambroxol HCl and salbutamol sulphate from tablet formulation by RP-HPLC. *GSC Biological and Pharmaceutical Sciences*. 2020; 13(03): 197-202.
96. Padgilwar S, et al. Traditional uses, phytochemistry and pharmacology of *Oroxylum Indicum*: A Review. *International Journal of Pharmaceutical and Phytopharmacological Research*. 2014; 3(6): 483-486.
97. Wadekar AB, et al. Morphology, phytochemistry and pharmacological aspects of *Carica papaya*, an review. *GSC Biological and Pharmaceutical Sciences*. 2020; 14(03): 234-248.
98. Gudalwar BR, et al. *Allium sativum*, a potential phytopharmacological source of natural medicine for better health. *GSC Advanced Research and Reviews*. 2021; 06(03): 220–232.
99. Manwar JV, et al. Experimental design approach for chromatographic determination of ketorolac tromethamine from bulk drug and tablet formulation. *Global Journal of Pharmacy & Pharmaceutical Sciences*. 2017; 3(2): 38-47.
100. Malode GP, et al. Phytochemistry, pharmacology and botanical aspects of *Murraya Koenigii* in the search for molecules with bioactive potential - A review. *GSC Advanced Research and Reviews*. 2021; 06(03): 143–155.
101. Manmode RS, et al. Stability indicating HPLC method for simultaneous determination of methocarbamol and nimesulide from tablet matrix. *Der Chemica Sinica*. 2011; 2(4): 81-85.
102. Bagade SB, et al. Simultaneous high performance thin layer chromatographic estimation of methocarbamol and nimesulide in combined dose tablet. *Journal of Pharmaceutical Research*. 2006; 5(4): 137-140.
103. Gulhane CA, et al. UV- Visible Spectrophotometric estimation of azithromycin and cefixime from tablet formulation by area under curve method. *World Journal of Pharmaceutical Sciences*. 2021; 9(6): 163-168.
104. Bijewar AH, et al. Overture in development, properties and clinical aspects of biosurfactants: A review. *International Journal of Medical, Pharmaceutical and Biological Sciences*. 2021; 1(1): 1-12.

105. Nikhare AM, et al. Morphological, phytochemical and pharmacological aspects of *Sygium cumini*. International Journal of Medical, Pharmaceutical and Biological Sciences. 2021; 1(1): 1-11.
106. Deshmukh SS, et al. Recent developments in niosomes, a smarter vesicular drug delivery system. International Journal of Medical, Pharmaceutical and Biological Sciences. 2021; 1(2):1.
107. Kukade SS, et al. Ethosomes: A revolutionary trend in lipid based vesicles and particulate carriers for transdermal delivery of drugs. International Journal of Medical, Pharmaceutical and Biological Sciences. 2021; 1(2):1.
108. Malode LL, et al. Potential of medicinal plants in management of diabetes: An updates. GSC Advanced Research and Reviews. 2021; 08(01): 149-159.
109. Manwar JV, et al. Comparative antioxidant potential of *Withania somnifera* based herbal formulation prepared by traditional and non-traditional fermentation processes. Integr Med Res. 2013; 2: 56-61.
110. Manwar JV, et al. Determination of withanolides from root and herbal formulation of *Withania somnifera* by HPLC coupled with DAD and ELSD detector. Der Pharmacia Sinica. 2012; 3 (1): 41-46.
111. Manwar JV, et al. Rapid RP-HPLC method for estimation of zidovudine from tablet dosage form. Der Chemica Sinica. 2011; 2(5): 152-156.
112. Panchale WA, et al. Microbeads: Generation, threat to biological and ecological systems and use of natural alternatives. International Journal of Medical, Pharmaceutical and Biological Sciences. 2021;1(2)1-10.
113. Patinge PA, et al. Ethnomedicinal, phytochemical and cosmeceutical updates on beauty plants from Indian origin. International Journal of Medical, Pharmaceutical and Biological Sciences. 2021;1(2):1-20.
114. Motule AS, Dongare PN, MP More, MG Nimbawar, SS Mankar, JV Manwar, RL Bakal. Progress in development of herbal cosmeceuticals: An current status and prospects. International Journal of Medical, Pharmaceutical and Biological Sciences. 2021; 1(1):1-9.
115. Bartere SA, Malode LL, Malode GP, Nimbawar MG, Gulhane CA, Manwar JV, Bakal RL. Exploring the potential of herbal drugs for the treatment of hair loss. GSC Biological and Pharmaceutical Sciences. 2021; 16(02):212-223.